P@LLINIS

12.2022

A NOT-FOR-PROFIT NGO REGISTERED UNDER FRENCH LAW, POLLINIS IS FUNDED EXCLUSIVELY BY DONATIONS FROM INDIVIDUALS TO PROTECT WILD AND HONEY BEES, AND TO PROMOTE SUSTAINABLE AGRICULTURE IN ORDER TO HELP PRESERVE POLLINATORS.

INTERNATIONAL APPEAL DANGEROUS BIOTECHNOLOGIES PUT POLLINATORS AT RISK AND THREATEN NATURE'S CONTRIBUTION TO PEOPLE

THIS IS AN APPEAL TO PROTECT INSECT POLLINATORS FROM THE POTENTIAL NEGATIVE EFFECTS OF BIOTECHNOLOGIES. IT IS AN INITIATIVE OF THE FRENCH NON-PROFIT ORGANISATION, POLLINIS, WHICH ACTS FOR THE PROTECTION OF POLLINATORS. THIS DOCUMENT WAS SIGNED BY KEY SCIENTISTS IN THE FIELDS OF MOLECULAR BIOLOGY, GENETICS, POLLINATOR ECOLOGY, AGROECOLOGY, KEY POLICY EXPERTS IN POLLINATOR PROTECTION, CONSERVATION, BEEKEEPING AND ENVIRONMENTAL PROTECTION, AND ORGANISATIONS.

KEY POINTS

- Pollinating insects are essential to biodiversity, ecosystem functions, and increase crop yield. In order to reverse their decline, we must provide them with a safe habitat within working landscapes where farming, ranching, and forestry take place.
- The release of organisms, products or components obtained through genetic biotechnologies, such as gene-silencing molecules (e.g. RNAi-based pesticides) and gene drive organisms (GDOs) could amplify the current stressors pollinators are already experiencing. To date, limited research has been conducted to understand the risks and impacts on pollinators of such a release.
- It is not possible to provide robust and reliable risk assessments to ensure that pollinators' decline will not be further precipitated by the release of these biotechnologies. Therefore, the signatories of this Appeal call for a strict application of the UN Precautionary Principle.
- We stress other ways to produce food based on biodiversity which are scientifically proven to achieve high yields and excellent nutrition quality, while not damaging the environment nor having the risks associated to the deployment of organisms through biotechnology in the environment.
- This Appeal, signed by prominent scientists, policy experts and organisations, calls upon the Parties and Signatories to the UN Convention on Biological Diversity to oppose the deployment in nature of genetic biotechnologies at international, regional and national levels.

APPEAL

We are appealing to Parties and Signatories to the UN Convention on Biological Diversity to oppose - at international, regional and national levels - the release of organisms, products and components obtained through genetic biotechnologies, including synthetic biology (1), genetic modification and genetic engineering, such as gene-silencing molecules (e.g. RNAi-based pesticides) and gene drive organisms (GDOs), in native habitats within both protected and working landscapes - that support human needs through farming, ranching, and forestry (2).

Such biotechnologies may harm insect pollinator populations and precipitate their ongoing decline. Despite urgent and documented warnings from the scientific community, the potential negative effects on pollinators, food webs and ecosystems of such a release in nature, remain understudied (3-5). We are therefore calling for a strict application of the UN Precautionary Principle (6), and to refrain from any releases until there is proof that there will be no negative impacts of direct or indirect effects of the application of these new genetic biotechnologies, their products, organisms and components.

"Pollinators have serviced the plants that they visit for at least 170 million years, since the mid-Mesozoic, and conceivably for far longer. Over that period the relative importance of different groups of pollinators has waxed and waned, while overall diversity has increased in parallel with flowering plants until, at the present time, there could be as many as 350 000 described species of pollinators (and many more awaiting scientific discovery). The relative importance of different taxonomic groups (from the levels of genus to order) varies biogeographically, but overall it is clear that diversity is important and loss of species (at whatever geographical scale) should be avoided" (p. 370) (7).

POLLINATORS NEED SAFE WORKING LANDSCAPES

Despite the challenges to estimate pollinator diversity, the most recent exhaustive report estimates that approximately 350 000 species of insects visit flowers and participate in pollination (7). Major groups are Lepidoptera (e.g. butterflies, moths), Hymenoptera (e.g. bees, bumblebees, wasps) and Diptera (e.g. flies, hoverflies). All these groups are currently facing a worldwide decline of their diversity and abundance (8-13), with an increasing number of species being found on the IUCN Red List as either Data Deficient (18.1 percent), Extinct (1 percent), Critically Endangered (3.1 percent), Endangered (9.1 percent), Vulnerable (11.4 percent) or Near Threatened (5.8 percent) (14).

This critical decline is due to several reasons, including conventional agriculture intensification, climate change, synthetic pesticides, pollution, pathogens, separately or in combination (15-18). These multiple anthropogenic pressures are linked to shifts in pollinator abundance and richness (19, 20). We are losing invaluable legacies from millenaries of evolution and plant-insect interactions (8, 21, 22), overall genetic diversity is impoverished, entire food webs are endangered as key species (23, 24) disappear and ecosystems might lose the fragile balance of which we know very little. As most flowering plants on Earth depend on insects to reproduce (25), plant diversity is now at risk worldwide (26, 27): with one plant out of five facing extinction (28). There is also rising concern about food production:

APPEAL

76 percent of the world's most important food crops (87 out of 115), including coffee, avocado and chocolate, require pollination by insects (29). Extensive research shows that increasing pollinator abundance and diversity increase crop yields (30-35). We need pollinators living and feeding in working landscapes, for sustainable agricultural production (30, 36-41). Hence, these working landscapes must be safe habitats for pollinators.

Since 1999, when the 'essential role of pollinators in sustainable agriculture and ecosystems' was internationally recognised in the "São Paolo Declaration of Pollinators" (42) and followed in 2000 by decision V/5, section II at COP-5 of the UN Convention on Biological Diversity (43), insect pollinators have been protected under numerous international agreements (26, 44). The role of pollinators in ecosystems will be an especially relevant topic at the COP15 in Montreal in December 2022 where Parties, government representatives, organisations, and indigenous people and local communities (IPLC) will gather to negotiate the Post-2020 Global Biodiversity Framework (GBF). Decisions that will be negotiated on some targets, specifically those addressing natural habitat, pollution, pesticide use and synthetic biology,¹ will have a direct impact on pollinators and the conditions of their survival.

Indeed, it is envisaged to open the way for potential release of organisms or products obtained through genetic biotechnologies. Agricultural applications include directly modifying insect genomes, or interfering with their gene expression, in order to change their behaviour or to make them extinct. All these applications, directly in native habitats within working landscapes, carry understudied risks which could accelerate the decline of pollinator populations and put entire food webs at risk.²

GENE DRIVE ORGANISMS (GDOs): MODIFYING INSECTS TO CHANGE THEIR BEHAVIOUR OR TO BECOME EXTINCT

Gene drive organisms are designed to spread engineered traits rapidly through populations. They are created with tools such as the CRISPR/Cas9 genome editing tool, which enables genes to be inserted, replaced, disrupted or deleted from DNA sequences. Gene drive (GD) systems are designed to override the rules of inheritance and force the spread of a trait to the next generation. Gene drive technologies aim not only to pass on an inserted or altered trait, but, additionally, to pass on the actual GD mechanism, including the "genetic scissor". The altered and added traits, as well as the genes encoded for the genome editing machinery, are then passed onto ALL offspring, causing the engineered genes as well as the GD mechanism with its genome editing processes to propagate fully through each generation, potentially in perpetuity (3).

^{1.} Targets 2, 3, 4, 7, 9, 10 and 17.

^{2.} Based on recent studies, understanding the complexity of interactions between and among organisms and plants suggest that the ecosystem is made up of many parts and pieces living together; they are known as larger units called holobionts or hologenomes, taking into account that all species in the same habitat interact and influence each other. Rosenberg et al. (2016) defines "holobiont" to include all animals and plants and introduced the term "hologenome" to describe the sum of the genetic information of the host and its symbiotic microorganisms" (pg. 1). They write: "The hologenome concept of evolution postulates that the holobiont (host plus symbionts) with its hologenome (host genome plus microbiome) is a level of selection in evolution. Multicellular organisms can no longer be considered individuals by the classical definitions of the term" (pg.1) [45].

APPEAL

A recent publication reported thirty-two insect targets, including twenty-one agricultural pests, from six different orders proposed or under GD technology development (45). For example, research has been undertaken to insert self-extinguishing genes in the spotted wing drosophila *(Drosophila suzukii)* (46), to target spermatogenesis of the common wasp *(Vespula vulgaris)* (47), and to remove olfactory functions of the noctuid moth *(Spodoptera littoralis)* (48) and the gypsy moth *(Lymantria dispar)* (49). Beyond these experiments, a number of companies have filed patent applications describing gene drive use in agriculture, including targeting hundreds of agricultural pests, in particular, WO 2017/049266 A2 (50) which consists of applying CRISPR-Cas9 gene drives on over three hundred agricultural pests (46, 50).

Gene drive organisms are expressly designed to spread, to create large-scale changes in natural populations and thus to transform entire ecosystems (51). Esvelt & Gemmell (2017) note that creating a standard, self-propagating CRISPRbased GD system is "equivalent to creating a new, highly invasive species" which can spread to any ecosystem in which it is viable, "possibly causing ecological change" (p.2) (51).

As synthetic GD uses the CRISPR gene modifying system, which has been observed to create unexpected 'off-target' effects (52-54). There is good reason to be concerned about unanticipated changes and mutations (55-58) that may recur with every generation as the CRISPR system is continually re-developed, not only within the lab but also in nature (3, 59).

It is possible that GD organisms could pass on engineered genes to closelyrelated species (3, 60) like insect pollinators by the vertical spread of genes via gene flow.³ They could also affect other non-target species via horizontal gene transfer (4). Limited studies have investigated these key issues (62), and the monitoring of these phenomena in the environment would be impossible (63).

Researchers have also raised concerns about transboundary contamination of agricultural systems related to the release of genetically modified insects as part of pest control strategies (64, 65). The release of such genetically modified insects in crop fields could irreversibly change the genetic makeup of managed (e.g. commercial honey bees and bumblebees) and wild insect populations, including non-target insects which are useful to industrial agriculture. Based on the GD organisms traits of forced spreadability and the fact that genetic modification processes continue to be active within them (due to the GD mechanisms engineered into them), a reliable risk assessment is not possible (66). As most applications are still in the stage of mathematical modeling, any release would be premature and would put entire ecosystems at risk.

^{3. «}Altered DNA could be transferred from organisms resulting from synthetic biology techniques to other organisms, either by sexual or horizontal gene flow/transfer» (p. 33) (61).

RNA-BASED TECHNOLOGIES: INTERFERING WITH THE GENE EXPRESSION OF INSECTS

Other technologies for environment-wide application include gene silencing molecules such as double stranded RNAs (dsRNAs), which are designed to fight crop pests or pathogens. They rely on sequence homology to target specific gene sequences, and use RNA interference mechanisms to silence genes responsible for vital functions in targeted insects, causing them to die. They can be delivered to crop pests through the mediation of genetically modified plants, bacteria, viruses, or directly applied as sprays (67).

Some dsRNA-based technologies are undergoing processes of approval, and some have already been approved by various national bodies for food, feed or cultivation purposes in many parts of the world (68-70). These issues must therefore be urgently addressed at the international level.

Many arthropod species share gene similarities, especially those belonging to the same taxonomic groups. Research has reported that a gene that is silenced and thus turns lethal for one species can also be lethal for another species (71). If two genes from two different species have a strong similarity, then there is a high probability that these two genes (of the same function) from two different species would be silenced by the same dsRNA (72).

There is limited understanding of how widespread the genetic similarities are among the different species. The current lack of independent research on non-target effects and homologous gene silencing needs to be addressed in order to assess the real danger for pollinators (73) and non-target species that live and feed in working landscapes.

Plans are also underway for genetically modified gut microbiota to deliver continuous dsRNA to honey bees in order to resist pesticides (74), parasites (75) or viruses (76). Whilst direct consequences of such microbial changes are not yet understood, it is also unclear whether contamination by genetically modified gut microorganisms of other species may occur through the pollination of common flowers, or whether or not this contamination may occur in honey products. Therefore, more research is necessary to be able to assess the direct and indirect effects of these biotechnologies applied to insect species, including pollinators.

REFRAMING THE DISCUSSION: A CALL FOR SOLUTIONS THAT RESPECT THE INTEGRITY OF ECOSYSTEMS

It is currently impossible to understand all the complex connections between and among species. Ecosystems are made up of multiple systems interacting with each other, on which scientists are continuing to make new discoveries and gain further understanding.⁴ The potential effects of applying genetic biotechnologies to open ecosystems could thus include dramatic changes in ecological networks structures and functions that could be disastrous for biodiversity.

It is clear that the current state of scientific research and knowledge is not able to provide a reliable and robust risk assessment to understand the effects of many new genetic biotechnologies and their applications on ecosystems and pollinators. Pollinating insects are already facing an alarming decline due to external stressors, adding hazardous and unassessed genetic biotechnologies to this fatal mix will aggravate the stress on pollinators and may precipitate their extinction.

We thus warn against the release of these genetic biotechnologies on pollinators as implications could be catastrophic. Our generation has a responsibility to pass on resilient and life-sustaining ecosystems, which include protected areas and native habitats in working landscapes (2). In order to create sustainable pathways toward secure food supplies, we need to rely on nature's contribution to people.⁵ It is vital to encourage ecological intensification for the improvement of crop yield (30, 77), rather than use genetic biotechnologies that may put entire ecosystems at risk.

Recent ecological research suggests that it is possible that the mutation of a single gene could potentially alter the structure and function of an ecosystem. For example, see Barbour, M., D. Kliebenstein and J. Bascompte (2022). «A keystone gene underlies the persistence of an experimental food web.» Science 376(6588): 70-73.
Indeed, evidence shows that enhancing pollinator abundance and diversity could close yield gaps by a median of 24 per cent. See Garibaldi, L., L. Carvalheiro, B. Vaissière, B. Gemmill-Herren, J. Hipólito, B. Freitas, H. Ngo, N. Azzu, A. Sáez, J. Åström, J. An, B. Blochtein, D. Buchori, F. Chamorro García, F. da Silva, K. Devkota, M. de Fátima Ribeiro, L. Freitas, M. Gaglianone, M. Goos, M. Irshad, M. Kasina, A. Pacheco Filho, L. Piedade Kill, P. Kwapong, G. Nates Parra, C. Pires, V. Pires, R. Rawal, A. Rizali, A. Saraiva, R. Veldtman, B. Viana, W. S and H. Zhang (2016). «Mutually beneficial pollinator diversity and crop yield outcomes in small and large farms.» Science 351(6217): 388-391.

Note: The signatories to this statement are signing in an individual capacity and not as representatives of their respective organisations.

Dr. Lucas A. Garibaldi

Director - IRNAD, Profesor - UNRN, Investigador Principal - CONICET (Argentina)

Dr Valeria Malagnini Researcher- Fondazione Edmund Mach (Italy)

Professor Antonio Felicioli Professor of biochemistry and professor of apidology- Pisa University (Italy)

Dr Paolo Fontana Researcher/ Entomologist - Edmund Mach Foundation (Italy)

Dr Jeff Pettis President of Apimondia - (USA)

Professor Thomas Dyer Seeley

Professor of Neurobiology and Behavior - Dept. of Neurobiology and Behavior, Cornell University (USA)

Christine von Weizsaecker President of Ecoropa - (Germany)

Professor Dave Goulson

Professor Biology and specialist in insect ecology - University of Sussex (UK)

Dr Benoît Geslin Associate professor- IMBE (France)

Dr Gérard Arnold Researcher - CNRS (France)

Dr Angelika Hilbeck Senior Researcher & Lecturer - ETH Zurich and ENSSER (Switzerland)

Dr Benjamin A. Woodcock Researcher Ecological Entomologist - (UK)

Dr Lionel Garnery Assistant professor Population geneticist - University Paris Saclay (France)

Dr Lanka Horstink Researcher - Institute of Social Sciences, sociology and political economy (Portugal)

Professor Mario Colombo Entomologist - Università Statale Milano (Italy)

Professor Marco Alberto Bologna Professor of Zoology and Biogeography- Department of Sciences, University Roma Tre (Italy)

Dr Bernard Vaissière Research Scientist at INRAE - (France)

Professor Paola Ferrazzi Entomologist, apidologist, zoologist - (Italy)

Ali Tapsoba Researcher at Terre à Vie - (Burkina Faso)

Dr Pablo Cavigliasso Researcher - Instituto Nacional de Tecnología Agropecuaria (Argentina)

Dr Fani Hatjina Researcher/ President of Bee Health Commission Apimondia - (Greece)

Dr Christos Astaras Wildlife Researcher at Forest Research Institute, ELGO-DIMITRA - (Greece)

Giovanni Timossi Researcher and Entomologist - (Italy)

Dr Lemeur Researcher at CNRS - (France)

François Warlop Agronomist (organic farming) - (France)

Professor Jean-Pierre Sarthou Professor of Agroecology- University of Toulouse- (France)

Dr Bettina Maccagnani Researcher on Biomonitoring programs through Honeybees funded by Automobili Lamborghini - (Italy)

Professor Bruno Massa Professor - University of Palermo (Italy)

Dr Nicola Palmieri Naturalist - (Italy)

Giuseppe Basso Politecnico Torino - (Italy)

Ilaria Negri Researcher Entomologist - (Italy)

Claudio Porrini University/technician - (Italy)

Martino Bertinotti Mechanical Engineer - (Italy)

Dr Andrea Spigolon Archaeologist - (Italy)

Stefano Fantini Entomology tutor - UNIBO : University of Bologna (Italy)

Professor Antonio De Cristofaro Entomologist - University of Molise (Italy)

Dr Carla Marangoni Curator - Museum of Zoology (Italy)

Dr Andrea Corso Zoologist - (Italy)

María Pilar Giovanetti Biologist - Instituto de Investigaciones en Recursos Naturales, Agroecologia y Desarrollo Rural (IRNAD-UNRN) (Argentina)

Fernanda Santibañez Doctoral student on Pollination - (Argentina)

Dr Anahí R. Fernandez Biologist - (Argentina)

Dr Paula Zermoglio Researcher in biology - Universidad Nacional de Río Negro - CONICET (Argentina)

Micaela Gambino IRNAD-University of Rio Negro - (Argentina)

Nicolas Laarman Director General – POLLINIS (France)

Romina Baroni Beekeeper - (Italy)

Dr Giorgio Galleano Journalist at Rai - (Italy)

Renato Galli Beekeeper La Risorgiva - (Italy)

Giuseppe Monaco Beekeeper - Apis Puglia APS (Italy)

Rachele Spezia Member of Federazione Apicoltori Italiani - (Italy)

Dr Danny Cliceri Science Coordinator - Resilient Bee Project (Italy)

Dr Giacomo Ciriello Beekeeper and Economist - (UK)

Dr Roberto Conte Trainer Health & Science - (Italy)

Dr Flavia Renzi Veterinary - (Italy)

Dr AnnaChiara Contri Veterinary and Beekeeper - (Italy)

Dr Laura Sommariva Journalist- (Italy)

Adam Breasley Biotechnology, human rights -(Australia)

Anne Petermann Executive Director of Global Justice Ecology Project - (USA)

Professor Guiomar Nates Parra Researcher - Universidad Nacional de Colombia (Colombia)

Barbara Pilz Campaign Manager at Save our Seeds - (Germany)

Ana Di Pangracio Law - (Argentina)

Dr Mariangela Fotelli Researcher (Forest ecophysiologist) - (Greece)

Dr Evangelos Hatzigiannakis

Research Director Agriculture - Soil and Water Resources Institute Hellenic Agricultural Organisation - (Greece)

Professor Aleksandar Uzunov Professor of honey bee biology and beekeeping - Ss Cyril and Methodius University in Skopje - (Macedonia)

Dr Grigorios Krey

Hellenic Agricultural Organization-Fisheries Research Institute/Biochemist-Molecular biologist - (Greece)

Dr Victor Kavvadias

Researcher in soil, Hellenic Agricultural Organization DIMITRA - (Greece)

Dalila Di Criscio

PhD student - Università degli studi del Molise (Italy)

Dr Georgios Tsoktouridis

Researcher - Institute of Plant Breeding and Phylogenetic Resources - (Greece)

Dr Eleni

Researcher in biology - (Greece)

Dr Thomas Sotiropoulos Researcher at Elgo Dimitra - (Greece)

Dr Dimitris Fotakis

Researcher at the Forest Research Institute - (Greece)

June Rebekka Bresson

Campaigner, MSc Integrated Food Studies - NOAH - Friends of the Earth Denmark (Denmark)

Dr Leonidas Researcher Apiculture Department Of Institute Animal Science - (Greece)

Aikaterini Karatasou

Veterinarian - Federation Of Greek Beekeepers Associations - Advisor - (Greece)

Naomi Kosmehl

Save Our Seeds / Gene Drives - (Germany)

Dr Catherine Wattiez

GMOs and pesticides campaigner at Nature et Progrès Belgique - (Belgium)

Dr Louise Vandelac

Professor and Researcher Sociology and Institute for environmental sciences - Université du Québec à Montréal (Canada)

Diederick Sprangers

Biochemist, and Board member of Genethics Fdn - (Netherlands)

Dr Mudssar Ali

Assistant Professor at MNS University of Agriculture of Multan and Pollinator Ecologist (Pakistan)

Ratia Gilles

International consultant at Apiservices - (France)

Professor Andreas Thrasyvoulou Professor of Apiculture - Aristotle Univesity Thessaloniki Greece (Greece)

Dr Savas Kazantzidis

Researcher at Forest Research Institute - (Greece)

Dr Giovanni Formato

Researcher in pathologies of the honey bee - Head of Apiculture Laboratory Honey bee pathologist (Italy)

Dr Peter

Researcher in the Slovenian Beekeepers Association - (Slovenia)

Dr Constantine Iliopoulos

Institute Director and Senior Researcher - Agricultural Economics Research Institute (AGRERI), Hellenic Agricultural Organization DEMETER (Greece)

Dr Cristina Mateescu

Senior Researcher in Biochemistry - (Romania)

Professor Giuseppe Longo

Directeur de recherches emeritus, formerly informatics, now philosophy of sciences-CNRS (France)

Dr Robert Chlebo

Professor of apiculture - Slovak University of Agriculture in Nitra (Slovakia)

Dr Michelle Leemans Post-doc - UPEC (France)

Dr Caro Gael

Researcher - University of Lorraine (France)

Dr Fabrice Requier

Researcher in Ecology - Laboratory EGCE and CNRS-IRD-Université Paris-Saclay (France)

Peter Sudovský

Citizens initiative Slovakia without GMO - (Slovakia)

Akiko Frid

GMO-Free regions - (Sweden)

Professor Erik Millstone

Professor Emeritus of Science Policy - Science Policy Research Unit, University of Sussex (UK)

Dr Nathalie Escaravage Researcher - Toulouse University (France)

Professor François Pompanon Prof. in Evolutionary Biology - University Grenoble Alpes (France)

Souparna Lahiri Climate and Biodiversity Policy Advisor - Global Forest Coalition (India)

Professor/Dr Johann Zaller

Professor of Ecology - University of Natural Resources and Life Sciences Vienna (BOKU) (Austria)

Dana Perls

Senior Food and Agriculture Program Manager - Friends of the Earth, U.S.; specialty: Emerging Technologies (USA)

Dr André Pornon

Researcher - Laboratoire Évolution et Diversité Biologique Université Paul Sabatier Toulouse (France)

Dr Schatz Bertrand

Researcher- CNRS (France)

Dr Jeavons Emma

Post doc and Ecologist specialized in agricultural ecosystems - INRAE (France)

Professor Hautekèete

Professor in Ecology & plant pollinator interactions- University of Lille (France)

Friedrich Wulf

International Biodiversity Campaigner at Friends of the Earth Europe - (Switzerland)

Professor Polyxeni Nicolopoulou Stamati

Professor of Environmental Pathology MD. PhD. - Medical School University of Athens (Greece)

Fabian Holzheid

Political director of Umweltinstitut München e.V. - (Germany)

Barbara Ntambirweki Researcher at ETC Group - (Uganda)

Tom Wakeford Europe Director ETC group

ORGANISATIONS

Vigilance OGM Save Our Seed (SOS Group) African Center for Biodiversity Friends of the Earth US ETC Group



1. Secretariat of the Convention on Biological Diversity (CBD). CBD Technical Series No. 100. Synthetic Biology. Montreal: CBD,; April 2022.

2. Garibaldi L, Oddi J, Miguez F, Bartomeus I, Orr M, Jobbágy E, et al. Working landscapes need at least 20% native habitat. Conservation Letters. 2020;14(2):e12773.

3. European Network of Scientists for Social and Environmental Responsibility, Critical Scientists Switzerland, Vereinigung Deutscher Wissenschaftler. Gene Drives. A report on their science, applications, social aspects, ethics and regulations. 2019.

4. National Academies of Sciences Engineering and Medicine. Gene Drives on the Horizon: Advancing Science, Navigating Uncertainty and Aligning Research with Public Values. Washington DC: The National Academies Press,; 2016.

5. Eckerstorfer M, Dolezel M, Greiter A, Miklau M, Heissenberger A, Steinbrecher R. Risk Assessment of Plants developed by new Genetic Modification Techniques (nGMs) Biosafety Considerations for Plants developed by Genome Editing and other new Genetic Modification... Bonn, Germany: BfN. Federal Agency for Nature Conservation,; 2020. Contract No.: BfN-Skripten 592.

6. United Nations General Assembly. Report of the United Nations Conference on Environment and Development. 1992. Contract No.: A/CONF.151/26 (Vol. I).

7. Ollerton J. Pollinator Diversity: Distribution, Ecological Function, and Conservation. Annual Reviews. 2017;48:353-76.

Zattara E, Marcelo A. Worldwide occurrence records suggest a global decline in bee species richness. One Earth. 2021;4(1):114 - 23.
Nieto A, Roberts S, Kemp J, Rasmont P, Kuhlmann M, García Criado M, et al. European Red List of Bees. Luxembourg: European Commission: 2014.

10. Hallmann C, Sorg M, Jongejans E, Siepel H, Hofland N, Schwan H, et al. More than 75 percent decline over 27 years in total flying insect biomass in protected areas. PLosOne. 2017.

11. Sánchez-Bayo F, Wyckhuys K. Worldwide decline of the entomofauna: A review of its drivers. Biological Conservation. 2019;232[April]:8-27.

12. Potts S, Biesmeijer J, Kremen C, Neumann P, Schweiger O, Kunin W. Global pollinator declines: trends, impacts and driver. Trends in Ecology and Evolution. 2010;25(6):345–53.

Van Swaay C, Cuttelod A, Collins S, Maes D, López Munguira M, Šaši M, et al. Red List of Butterfies. European Luxembourg; 2010.
IUCN. IUCN's Red List of Threatened Species 2022 [Available from: <u>https://www.iucnredlist.org/about</u>.

15. González-Varo J, Biesmeijer J, Bommarco R, Potts S, Schweiger O, Smith H, et al. Combined effects of global change pressures on animal-mediated pollination. Trends in Ecology & Evolution, 28, 524–534. Trends in Ecology & Evolution. 2013;28(9):524–30.

16. Vanbergen A, The Insect Pollinators Initiative. Threats to an ecosystem service: pressures on pollinators. Frontiers in Ecology and the Environment. 2013;11(251–259).

17. IPBES. The assessment report of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services on pollinators, pollination and food production. Bonn, Germany: Secretariat of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services, ; 2016.

18. Dicks L, Breeze T, Ngo H, Senapathi D, An J, Aizen M, et al. A global-scale expert assessment of drivers and risks associated with pollinator decline. Nature ecology and evolution. 2021;5:1453-61.

19. Carvalheiro L, Kunin W, Keil P, Aguirre-Gutiérrez J, Ellis W, Fox R, et al. Species richness declines and biotic homogenisation have slowed down for NW-European pollinators and plants. Ecology Letters. 2013;16(7):870-8.

20. Senapathi D, Carvalheiro L, Biesmeijer J, Dodson C-A, Evans R, McKerchar M, et al. The impact of over 80 years of land cover changes on bee and wasp pollinator communities in England. Proceedings of the Royal Society B: Biological Sciences. 2015;282(20150294).

21. Kiester A, Lande R, Schemske D. Models of Coevolution and Speciation in Plants and Their Pollinators. The American Naturalist 1984;124[2]:220-43.

Hu S, Dilcher D, Jarzen D, Winship T. Early steps of angiosperm pollinator coevolution. Proc Natl Acad Sci U S A 2008;105(1):24-245.
Huaylla C, Nacif M, Coulin C, Kuperman M, Garibaldi L. Decoding information in multilayer ecological networks: The keystone species case, Volume 460, 2021, 109734, ISSN 0304-3800,. Ecological Modelling. 2021;460(109734).

24. Garibaldi A, Turner N. Cultural Keystone Species: Implications for Ecological Conservation and Restoration. Ecology and Society.9(3):Art. 1.

25. Ollerton J, Winfree R, Tarrant S. How many flowering plants are pollinated by animals? Oikos. 2011;120:320-6.

26. IPBES. Global assessment report on biodiversity and ecosystem services of the Intergovernmental Science-Policy Platform on Biodiversity and Ecosystem Services. Bonn, Germany: IPBES Secretariat; 2019.

27. Biesmeijer J, Roberts S, Reemer M, Ohlemüller R, Edwards M, Peeters T, et al. Parallel declines in pollinators and insect-pollinated plants in Britain and the Netherlands. Science. 2006;313(5785):351-4.

28. Vanbergen A. Causes and consequences of pollinator decline. Brussels: Belgium Science Policy Office (BELSPO); 2018.

29. Klein A, Vaissière, Cane J, Steffan-Dewenter I, Cunningham S, Kremen C, et al. Importance of pollinators in changing landscapes for world crops. Proceedings of the Royal Society B 2006;274:303-13.

30. Garibaldi L, Pérez-Méndez N, Garratt M, Gemmill-Herren B, Miguez F, Dicks L. Policies for Ecological Intensification of Crop Production. Trends in Ecology & Evolution. 2019;34(4):282-6.

31. Chagnon M, Gingras J, de Oliveira D. Complementary Aspects of Strawberry Pollination by Honey and IndigenQus Bees (Hymenoptera). Journal of Economic Entomology. 1993;86(2):416–20.

32. Fontaine C, Dajoz I, Meriguet J, Loreau M. Functional Diversity of Plant–Pollinator Interaction Webs Enhances the Persistence of Plant Communities. PLOS Biology. 2005;4(1):e1.

33. Hoehn P, Tscharntke T, Tylianakis J, Steffan-Dewenter I. Functional group diversity of bee pollinators increases crop yield Proc R Soc B. 2008;275(1648).

34. Garibaldi L, Steffan Dewenter I, Kremen C, Morales J, Bommarco R, Cunningham S, et al. Stability of pollination services decreases with isolation from natural areas despite honey bee visits. Ecology Letters. 2011;14:1062-72.

35. Bartomeus I, Park M, Gibbs J, Danforth B, Lakso A, Winfree R. Biodiversity ensures plant-pollinator phenological synchrony against climate change Ecology Letters. 2013;16:1331-8.

36. Potts S, Imperatriz-Fonseca V, Ngo H, Aizen M, Biesmeijer J, Breeze T, et al. Safeguarding pollinators and their values to human well-being. Nature. 2016;540:220-9.

37. Kevan P, Phillips T. The economic impacts of pollinator declines: an approach to assessing the consequences. Conservation Ecology 2001;5(1):8.

38. Smith M, Singh G, Mozaffarian D, Myers S. Effects of decreases of animal pollinators on human nutrition and global health: a modelling analysis. The Lancet. 2015;386(10007):P1964-72.

39. Stenchly K, Hansen M, Stein K, Buerkert A, Loewenstein W. Income vulnerability of west african farming households to losses in pollination services: a case study from Ouagadougou, Burkina Faso. Sustainability. 2018;10(4253).

40. Chaplin-Kramer R, Sharp R, Weil C, Bennett E, Pascual U, Arkema K, et al. Global modeling of nature's contributions to people. Science. 2019;366:255-8.

41. Aizen M, Aguiar S, Biesmeijer J, Garibaldi L, Inouye D, Jung C, et al. Global agricultural productivity is threatened by increasing pollinator dependence without a parallel increase in crop diversification. Global change biology. 2019;25(10):3516-27.

42. Dias B, Raw A, Imperatri-Fonseca V. International Pollinators Initiative: The São Paulo Declaration on Pollinators. Brasília: Brazilian Ministry of the Environment; 1999.

43. Convention on Biological Diversity. COP5 Decision V/5. Retired sections: paragraphs 1-2, 8, 20-21 and 28-29. Agricultural biological diversity: review of phase I of the programme of work and adoption of a multi-year work programme. 1999.

44. CBD. Report of the Subsidiary Body on Scientific, Technical and Technological Advice on its Twenty-Third Meeting. Montreal: CBD. 45. Wells M, Steinbrecher R. Current and proposed insect targets for gene drive development. A tabular overview. EcoNexus; 2022.

46. ETC Group. Forcing the Farm. How Gene Drive Organisms Could Entrench Industrial Agriculture and Threaten Food Sovereignty. 2018.

47. Lester P, Bulgarella M, Baty J, Dearden P, Guhlin J, Kean J. The potential for a CRISPR gene drive to eradicate or suppress globally invasive social wasps. Scientific Reports. 2020;10(12398).

48. Koutroumpa F, François M, de Cian A, Royer C, Concordet J, Jacquin-Joly E. Heritable Genome Editing with CRISPR/ Cas9 Induces Anosmia in a Crop Pest Moth. Scientific Reports. 2016;6(29620).

49. Esvelt K. Readings: Gene Drives And CRISPR Could Revolutionize Ecosystem Management Wyss Institute2014.

50. Bier E, Gantz V, Hedrick S, inventorsMethods for Autocatalytic Genome Editing and Neutralizing Autocatalytic Genome Editing and Compositions Thereof. USA2017.

51. Esvelt K, Gemmell N. Conservation demands safe gene drive. PLoS Biology. 2017;15.

52. Honeybee Genome Sequencing Consortium. Insights into social insects from the genome of the honeybee Apis mellifera. Nature. 2006;443[7114]:931-49.

53. Yang Q, Tae-Sung P, Bumkyu L, Myung-Ho L. Unusual Removal of T-DNA in T1 Progenies of Rice after Agrobacterium-mediated CRISPR/Cas9 Editing. Research Square. 2022.

54. Fu Y, Foden J, Khayter C, Maeder M, Reyon D, Joung J, et al. High-frequency off-target mutagenesis induced by CRISPR-Cas nucleases in human cells. Nat Biotechnol. 2013;31(9):822-6.

55. Gelinsky E, Hilbeck A. European Court of Justice ruling regarding new genetic engineering methods scientifically justified: a commentary on the biased reporting about the recent ruling. Environmental Sciences Europe. 2018;30(52).

56. Adikusuma F, Piltz S, Corbett M, Turvey M, McColl S, Helbig K, et al. Large deletions induced by Cas9 cleavage. Nature. 2018;560:8-9.

57. Kosicki M, Tomberg K, Bradley A. Repair of double-strand breaks induced by CRISPR-Cas9 leads to large deletions and complex rearrangements. Nature Biotechnology. 2018;36:765-71.

58. Steinbrecher R, Paul H. New Genetic Engineering Techniques: Precaution, Risk, and the Need to Develop Prior Societal Technology Assessment. Environment: Science and Policy for Sustainable Development. 2017;3(59):38–47.

59. Wells M, Steinbrecher R. Natural selfish genetic elements should not be defined as gene drives. PNAS. 2022;119(34): e2201142119.

60. Courtier-Orgogozo V, Danchin A, Gouyon P, Boëte C. Evaluating the Probability of CRISPR-based Gene Drive Contaminating Another Species. bioRxiv. 2019.

61. CBD. Synthetic Biology. Part I: Potential impacts of synthetic biology on biological diversity. Part II: Gaps and overlaps with the provisions of the convention and other agreements. Montreal; 2015. Contract No.: CBD Technical Series No. 82.

62. Dröge M, Pühler A, Selbitschka W. Horizontal gene transfer as a biosafety issue: a natural phenomenon of public concern. J Biotechnology. 1998;64(1):75-90.

63. Heinemann J, Traavik T. Problems in monitoring horizontal gene transfer in field trials of transgenic plants. Nat Biotechnol. 2004;22(9):1105-9.

64. Hayes K, Hosack G, Dana G, Foster S, Ford J, Thresher R, et al. Identifying and detecting potentially adverse ecological outcomes associated with the release of gene-drive modified organisms. Journal of Responsible Innovation. 2018;5(51):S139–S58.

65. Reeves R, Phillipson M. Mass Releases of Genetically Modified Insects in Area-Wide Pest Control Programs and Their Impact on Organic Farmers. Sustainability. 2017;9(59).

66. Sirinathsinghji E. Risk assessment challenges of synthetic gene drive organisms. Biosafety Information Centre; 2020.

67. Sirinathsinghji E, Klein K, Perls D. Gene-Silencing Pesticides. Risks and Concerns. Friends of the Earth USA; 2020.

68. Li X, Liu X, Lu W, Yin X, An S. Application progress of plant-mediated RNAi in pest control. Frontiers in Bioengineering and Biotechbology. 2022;10(963026).

69. OECD. BioTrack Product Database [Available from: https://biotrackproductdatabase.oecd.org/Product.aspx?id=MON-87411-9.

70. Jalaluddin N, Othman R, Harikrishna J. Global trends in research and commercialization of exogenous and endogenous RNAi technologies for crops. Critical Reviews in Biotechnology. 2019;39(1).

71. Mogilicheria K, Howell J, Reddy Palli S. Improving RNAi in the Brown Marmorated Stink Bug: Identification of target genes and reference genes for RT-qPCR. Scientific Reports. 2018;8(3720).

72. Chen J, Peng Y, Zhang H, Wang K, Zhao C, Zhu G, et al. Off-target effects of RNAi correlate with the mismatch rate between dsRNA and non-target mRNA. RNA Biol. 2021;18(11):1747-59.

73. Casacuberta J, Devos Y, du Jardin P, Ramon M, Vaucheret H, Nogué F. Biotechnological uses of RNAi in plants: risk assessment considerations. Trends Biotechnol. 2015;33(3):145-7.

74. Kovarik J, inventorMethod and System for Protecting Honey Bees, Bats and Butterflies From Neonicotinoid Pesticides. US2016. 75. In the pipeline: protecting the honeybee [press release]. 2019.

 Hunter W, Ellis J, Vanengelsdorp D, Hayes J, Westervelt D, Glick E, et al. Large-scale field application of RNAi technology reducing Israeli acute paralysis virus disease in honey bees (Apis mellifera, Hymenoptera: Apidae) . PLOS Pathogens. 2010;6(12).
Kovács-Hostyánszki A, Espíndola A, Vanbergen A, Settele J, Kremen C, Dicks L. Ecological intensification to mitigate impacts of conventional intensive land use on pollinators and pollination. Ecology Letters 2017;20(673–689).